## SULPHATE ESTERS OF HYDROXYCINNAMIC ACID-SUGAR DERIVATIVES FROM ADIANTUM CAPILLUS-VENERIS

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**Key Word Index**—Adiantum capillus-veneris; Adiantaceae; sulphate esters of 1-p-coumarylglucose, 1-caffeylglucose and 1-caffeylgalactose.

Abstract—Two new and two known sulphate esters of hydroxycinnamic acid-sugar derivatives have been isolated from the fronds of the fern *Adiantum capillus-veneris*. The new compounds have been shown to be 1-p-coumarylglucose 2-sulphate and 1-caffeylgalactose 6-sulphate by chemical and spectroscopic methods.

Previous work on the chemical constituents of the fern, Adiantum capillus-veneris L., has shown the presence of triterpenoids [7,9(11)-fernadiene, 7-fernene [1],  $3\alpha.4\alpha$ -epoxyfilicane [2],  $21\beta$ -hydroxy-29-nor-22-hopanone [3] and adiantone [4]], alicyclic acids [5] (quinic acid and shikimic acid) and of a flavonol glycoside (kaempferol 3,7-diglucoside [6]). This paper describes the identification of four sulphate esters of hydroxycinnamic acid-sugar derivatives.

From an ethanolic extract of fresh fronds of Adiantum capillus-veneris, two bands (B1 and B2) were isolated by prep. PC and their chromatographic and electrophoretic properties are presented in Table 1. B1 (colour reactions: colourless to blue in UV+ ammonia) was electrophoretically highly mobile towards the anode. Both total acid hydrolysis and controlled acid hydrolysis gave p-coumaric acid, Dglucose and sulphate. The UV spectral properties  $(\lambda_{\text{max}}^{\text{MeOH}} \text{ nm: } 230, 315; +AlCl_3, 231, 316; +NaOMe 240,$ 365) showed that the phenolic hydroxyl group of p-coumaric acid must be free (large bathochromic shift of the band at 315 nm in the presence of sodium mectoxide [7]).  $R_i$  values, colour reactions and UV spectral properties of B1 were in agreement with those of an authentic sample of the sulphate ester of 1-p-coumarylglucose from Adiantum pulverulentum [8]. Treatment with sulphatase gave 1-p-coumaryl-glucose [7] identified by UV spectroscopy, acid hydrolysis, alkaline hydrolysis, treatment with  $\beta$ -glucosidase and co-PC. Kuhn methylation followed by acid hydrolysis gave p-methoxycinnamic acid, 2,3,4-tri-O-methyl-D-glucose and 3,4,6-tri-O-methyl-D-glucose. These results show that B1 must be a mixture of 1-p-coumarylglucose 6-sulphate (1) and 1-p-coumarylglucose 2-sulphate (2). 1 may already have been found in plants since it was suggested [8] that sulphate is probably attached to the 6-hydroxy group of the glucose in the sulphate ester of 1-p-coumarylglucose isolated from Adiantum pulverulentum [8]. However, 2 is a new natural product.

Band B2 (colour reactions: blue to green in UV + ammonia) migrated towards the anode on electrophoretograms. Both total acid hydrolysis and controlled acid hydrolysis gave caffeic acid, D-glucose, D-galactose and sulphate. The UV spectral properties  $[\lambda_{\text{max}}^{\text{MEOH}}$  nm: 245, 300 (sh), 332; +NaOMe 268, 320 (sh), 372; +AlCl<sub>3</sub> 250, 300 (sh), 340; +AlCl<sub>3</sub>-HCl 250, 318 (sh), 330; NaOAc-H<sub>3</sub>BO<sub>3</sub> 258, 302 (sh), 344] showed that the O-diphenolic group of caffeic acid must be free. Enzymic hydrolysis with sulphatase gave a mixture of 1-caffeylglucose [7] and 1-caffeylgalactose which were identified by UV spectroscopy,

Table 1.  $R_f$  and electrophoretic mobility of bands isolated from Adiantum capillus-veneris

	$R_f(\times 100)$ in					Electronic materials
Band	BAW	BEW	15% HOAc	30% HOAc	H <sub>2</sub> O	— Electrophoretic mobility*
B <sub>1</sub>	45	53	72	85	82	12.3
$\mathbf{B}_2$	22	35	68	77	70	9.2

<sup>\*</sup>Relative to p-coumaric acid run at pH 4.5 (pyridine formate 0.05 M) at 16 V/cm for 3 hr on Whatman No. 1 paper.

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acid hydrolysis, alkaline hydrolysis, treatment with  $\beta$ -glucosidase and co-PC.  $R_t$  values, colour reactions and UV spectral properties of B2 were in agreement with those of an authentic sample of the sulphate ester of 1-caffeylglucose [8] from Pteridium aquilinum. Kuhn methylation followed by acid hydrolysis gave 3,4-dimethoxycinnamic acid, 2,4,6-tri-O-methyl-D-glucose and 2,3,4-tri-O-methyl-D-galactose. These results show that B2 must be a mixture of 1-caffeylglucose 3-sulphate (3) and 1-caffeylgalactose 6-sulphate (4). 3 has recently been found in the fern Ceterach officinarum Lam. et DC. [9]; 4 is a new natural product. This is also the first report of a sulphated cinnamic acid galactoside.

 $I R_1 = R_2 = R_3 = R_5 = H_1 R_4 = OH_1 R_6 = SO_3$ 

**2**  $R_1 = R_3 = R_5 = R_6 = H$ ;  $R_4 = OH$ ;  $R_2 = SO_3$ 

**3**  $R_2 = R_5 = R_6 = H$ ;  $R_1 = R_4 = OH$ ;  $R_5 = SO_3$ 

**4**  $R_2 = R_3 = R_4 = H$ ;  $R_1 = R_5 = OH$ ;  $R_6 = SO_3^{-1}$ 

Sulphated cinnamic acid glucosides were first discovered [8] in 1975 by Cooper-Driver and Swain who reported the presence of sulphate esters of 1-caffeyl-glucose and 1-p-coumarylglucose in Pteridium aquilinum and in 10 Adiantum species. Subsequently, 2-O-p-coumarylglucose 6-sulphate was isolated [9] from Asplenium fontanum Bernh var. obovatum and the 3- and 2-sulphates of 1-caffeylglucose were found in Ceterach officinarum [9]. The occurrence of sulphate esters of hydroxycinnamic acid-sugar derivatives is not restricted to lower plants since sulphate esters of caffeylglucose have been found in higher plants (e.g. in the grass Paspalum convexum [10]).

## EXPERIMENTAL

Plant material. Fronds of Adiantum capillus-veneris L. were collected in Catania.

Isolation. Fresh fronds (100 g) of A. capillus-veneris were homogenized and extracted ×3 with hot 95% EtOH. The combined extracts were filtered, concd. to small vol. in vacuo and re-filtered. Prep. PC on Whatmann 3MM paper in BAW afforded two bands (B1 and B2) which were eluted with 70% EtOH, concd. and rechromatographed in 15% HOAc; the bands, B1 (30 mg) and B2 (50 mg), were then purified by chromatography in BEW. Another anionic band was isolated but was not present in sufficient amount for analysis; electrophoretic and spectral properties as well as colour reactions suggested that this band may be a sulphate ester of a caffeic acid derivative.

Hydrolysis procedures. Controlled acid hydrolysis was carried out with 10% aq. HOAc (3.5 hr under reflux); total acid hydrolysis was carried out with 2 N HCl (2 hr at 100°); alkaline hydrolysis was carried out with 2 N NaOH at room temp. for 3 hr (under N<sub>2</sub>); enzyme hydrolysis with sulphatase (from Helix pomatia) was carried out in citrate-Pi buffer, pH 4.5 at 37° for 12 hr. D-Glucose and D-galactose were identified by co-PC (four solvents) and TLC on Si gel (n-BuOH-HOAc-Et<sub>2</sub>O-H<sub>2</sub>O, 9:6:3:1). Sulphate was identified by BaCl<sub>2</sub>.

Methylation. The bands were methylated with MeI in HCONMe<sub>2</sub> in the presence of AgO and the permethylated products were hydrolysed with 0.3 N HCI (4 hr under reflux). 3,4-Dimethoxycinnamic acid and p-methoxycinnamic acid were identified by PC [11] and TLC on Si gel; the partially methylated sugars were identified by PC according to Petek [12] and, after reduction (NaBH<sub>4</sub>) and acetylation (pyridine-Ac<sub>2</sub>()), by GC/MS as partially methylated alditol acetates (70 eV; stainless steel column packed with 3% ECNSS-M on Gas-Chrom Q 1.80 m × 1.8 mm, at 180° isothermally and N<sub>2</sub> at 25 ml/min; injector temp. 200°, FID temp. 200°).

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